



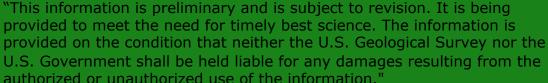
## eDNA Monitoring Research

- USGS round-robin assay evaluation
- USGS streamgage eDNA sampling

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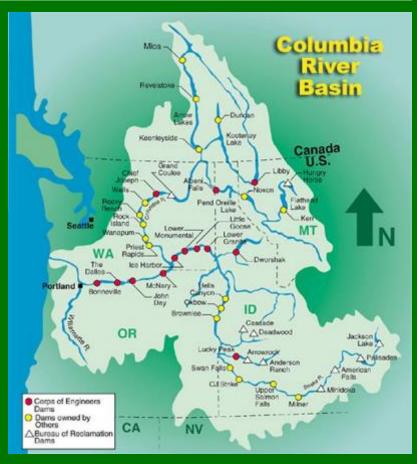




### Need

- 1. How to sample?
- 2. Where to sample?









# Round-robin evaluation of eDNA assays

#### eDNA is powerful early detection tool but...

#### Uncertainty associated with eDNA results

- Indirect method that detects DNA
  - DNA presence ≠ Species presence
- Often impossible to corroborate with direct methods
- Newer science with checkered early history





## Towards reducing uncertainty of eDNA results...

## Identify lab-based protocols that provide repeatable & reproducible results.

- Lab environment can be totally controlled
- Repeatable: same sample & conditions = same results
- Reproducible: same sample, different conditions same result
- Cornerstone good science & diagnosis
  - confidence that initial results not obtained by chance or errors.





## Approach, 5 assays & 4 labs

- Public, probe-based quantitative PCR assays
  - 1. DRE16S: zebra & quagga
  - 2. DRE2: zebra
  - 3. ZebCOI: zebra
  - 4. ZebCYT: zebra
  - 5. DREQM: quagga
- Four USGS labs
  - Boise, ID; Bozeman, MT; LaCrosse, WI; Seattle, WA





## **Approach**

#### With mussels

- Seneca Lake (ZM & QM)
- Lake Michigan (zm & QM)
- San Justo Res (ZM)
- Lake Mead (QM)



#### No mussels, spiked with ZM + QM DNA

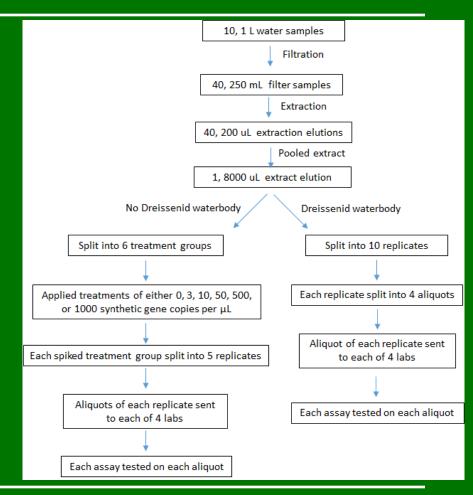
- Yellowstone River
- Jackson Lake
- Columbia River





## **Approach**

- Partners collected samples using SOP
- Double-blind
  - Sample prep
  - Sample analysis
- Identical samples
- 2832 samples, 8497 qPCR reps







## Repeatability ~ 90%

Likelihood that two identical samples analyzed under the same conditions (same assay or same lab) will yield same result

1 = results are the same

Assay	Mean	CI95
DRE16S	0.92	0.03
DREQM	0.93	0.02
DRE2	0.85	0.03
ZEBCOI	0.91	0.03
ZEBCYT	0.93	0.03
Lab	Mean	Cl95
1	0.93	0.03
2	0.92	0.03
3	0.92	0.03
4	0.86	0.04





### Reproducibility ~ 0.90

Likelihood that two identical samples analyzed under different conditions (assay or lab) will yield same result

• 1 = results are the same

	Mean	CI95	
Assays	0.92	0.01	
Labs	0.92	0.01	
Assay	DRE16S	ZEBCOI	ZEBCYT
DRE2	0.90	0.90	0.90
ZEBCYT	0.93	0.93	
ZEBCOI	0.93		
DREQM	0.95		
Lab	4	3	2
1	0.89	0.93	0.94
2	0.90	0.93	
3	0.87		





## **Conclusions & Next Steps**

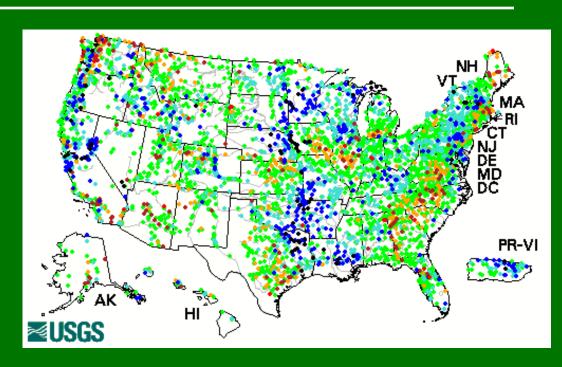
- High probability (~90%) that identical samples analyzed under the same or different assays or labs will yield the same result
- Dreissenid mussel eDNA: increase confidence / decrease uncertainty in results
- Benchmark for new assays and /or labs
- Next steps: evaluate eDNA field sampling protocols





## **USGS Streamgage eDNA sampling**

- 10,000+ USGS streamgages
- USGS hydro techs visit ~ monthly
- Incorporate eDNA sampling into visits
  - Cheap and effective way to help Safeguard the West?

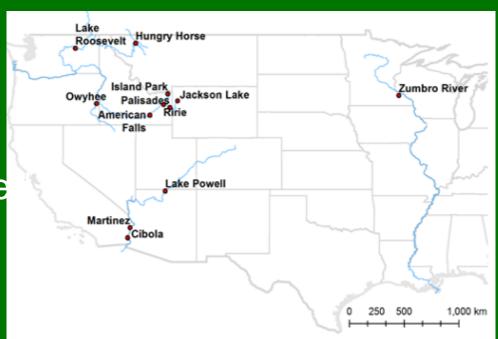






## **USGS** streamgages below reservoirs

- Reservoirs
  - invasion hotspots
  - critical infrastructure
- USGS gages often locate below reservoirs
- eDNA at gages
  - Mussel (-), vulnerable
  - Mussel (+)







## **Objectives**

- 1. eDNA surveillance for dreissenid mussels
- 2. Additional cost of incorporating eDNA into USGS streamgage program
- 3. Are streamgages adequately located for biosurveillance of upstream reservoirs?
  - What influences increase detection rates?





## **Approach**

- Hydro-techs trained in SOP
  - 3 10 water samples per visit
  - May September 2018
  - Shipped/analyzed at USGS LaCrosse, WI
- Target species
  - Dreissenid mussels
    - Likely all no-detects... hard to learn
  - Kokanee salmon & Yellow perch
    - Reservoir 'obligates' at varying abundance
    - Used these results to learn





#### Results

#### 1. Dreissenid mussel DNA

- Mussel pos. waters: all samples positive
- Mussel neg. waters: no sample met criteria for being scored as positive

#### 2. Additional costs:

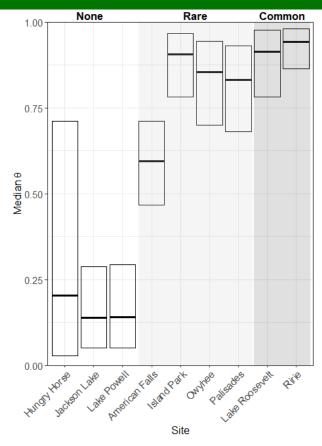
- \$400 500 for three samples at a streamgage
- Additional 2 hr of hydro-tech time
- 3. Streamgages adequately located
  - Kokanee and yellow perch DNA: yes



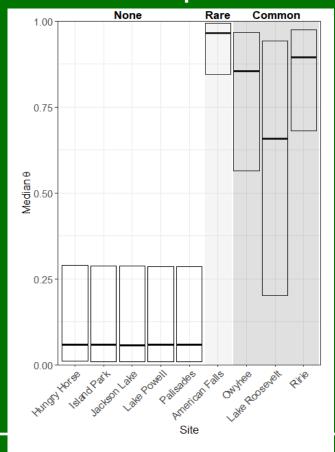


## Results: detection rate in 1 sample

#### Kokanee



#### Yellow perch







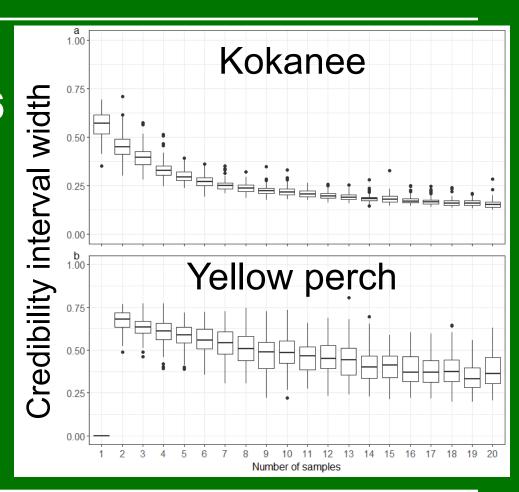
## Results: how many samples?

#### Kokanee

diminishing returns > 6

#### Yellow perch

- Less certain, but likely
  - > 10







#### Take homes

#### Streamgage eDNA sampling

- always detected mussel DNA when mussels are present
- sensitive even when fish at low abundance
- cost-effective

#### **Next steps**

- What are we missing by limiting to streamgage?
  - Send USGS hydro techs to reservoirs rather than, or in addition to, streamgage?



